

RESEARCH

Open Access



The causal relationship between human blood metabolites and the risk of visceral obesity: a mendelian randomization analysis

Zhaoxiang Wang¹ and Qichao Yang^{2,3*}

Abstract

Background We aimed to explore the causal relationship between blood metabolites and the risk of visceral obesity, as measured by visceral adipose tissue (VAT).

Methods Summary statistics for 486 blood metabolites and total, as well as sex-stratified, MRI-derived VAT measurements, adjusted for body mass index (BMI) and height, were collected from previous genome-wide association studies (GWAS). A two-sample Mendelian Randomization (MR) design was used. Comprehensive evaluation was further conducted, including sensitivity analysis, linkage disequilibrium score (LDSC) regression, Steiger test, and metabolic pathway analysis.

Results After multiple testing correction, arachidonate (20:4n6) has been implicated in VAT accumulation ($\beta=0.35$, 95%CI:0.18–0.52, $P<0.001$; FDR=0.025). Additionally, several blood metabolites were identified as potentially having causal relationship (FDR<0.10). Among them, lysine ($\beta=0.67$, 95%CI: 0.28–1.06, $P<0.001$; FDR=0.074), proline ($\beta=0.30$, 95%CI:0.13–0.48, $P<0.001$; FDR=0.082), valerate ($\beta=0.50$, 95%CI:0.23–0.78, $P<0.001$, FDR=0.091) are associated with an increased risk of VAT accumulation. On the other hand, glycine ($\beta=-0.21$, 95%CI: -0.33–0.09), $P<0.001$, FDR=0.076) have a protective effect against VAT accumulation. Most blood metabolites showed consistent trends between different sexes. Multivariable MR analysis demonstrated the effect of genetically predicted arachidonate (20:4n6) and proline on VAT remained after accounting for BMI and glycated hemoglobin (HbA1c). There is no evidence of heterogeneity, pleiotropy, and reverse causality.

Conclusion Our MR findings suggest that these metabolites may serve as biomarkers, as well as for future mechanistic exploration and drug target selection of visceral obesity.

Keywords Visceral obesity, Blood metabolites, GWAS, LDSC, Mendelian randomization analysis

*Correspondence:

Qichao Yang
yangqichao@wjrmmy.cn

¹Department of Endocrinology, Affiliated Kunshan Hospital of Jiangsu University, Kunshan, Jiangsu 215300, China

²Department of Endocrinology, Affiliated Wujin Hospital of Jiangsu University, Changzhou, Jiangsu 213017, China

³Wujin Clinical College of Xuzhou Medical University, Changzhou, Jiangsu 213017, China



© The Author(s) 2024. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

Introduction

Obesity is a major global health problem, affecting billions of people worldwide and showing significant heterogeneity [1, 2]. The distribution of adipose tissue was considered as an important factor in determining the health risks of obesity [1]. VAT, the intra-abdominal fat encapsulating internal organs, is metabolically active and potentially harmful, unlike subcutaneous adipose tissue (SAT) [2, 3]. Visceral obesity, characterized by excessive VAT accumulation, is significantly linked to numerous detrimental health conditions, including cardiovascular diseases, metabolic disorders, and cancers [2, 4, 5].

Metabolomics, a modern omics-based technology, has greatly enhanced our understanding of disease mechanisms by uncovering intermediate metabolites and altered metabolic pathways [6]. The analysis of extensive datasets generated by previous metabolomics studies has revealed the vital role of metabolic products, such as lipids, amino acids, and so on, in regulating energy metabolism, lipid synthesis, adipocyte differentiation, lipid oxidation, and insulin sensitivity [7]. Nevertheless, the precise association between blood metabolites and the distribution of adipose tissue remains elusive in the current scientific understanding. Thus, it is imperative to identify metabolites associated with adipose tissue, particularly VAT, to investigate the potential metabolic mechanisms of visceral obesity and develop targeted intervention measures.

Compared to conventional observational studies, the MR approach is a powerful method for obtaining robust evidence of causality utilizing genetic variants as instrumental variables (IVs) to address confounding [8]. In this study, we followed the STROBE-MR guidelines and conducted a two-sample MR analysis to investigate the potential causal association between human blood metabolites and VAT accumulation [9].

Methods

Study design

The MR analysis was conducted based on three crucial assumptions: (1) The IVs used in the analysis exhibited a strong association with blood metabolites. (2) The chosen IVs and potential confounding variables, which could impact blood metabolites and VAT, were not interrelated. (3) The IVs solely influenced VAT through their impact on blood metabolites.

Data sources

Blood metabolite genetic data were obtained from the metabolomics GWAS server (<http://metabolomics.helmholtz-muenchen.de/gwas/>), or the GWAS Catalog (<https://www.ebi.ac.uk/gwas/>) [10, 11]. Shin et al. performed a comprehensive GWAS of non-targeted metabolomics, identifying 486 human serum metabolites with

genetic influences [11]. A total of 7824 participants were enrolled from two European population cohorts: 1768 participants from the KORA F4 study in Germany and 6056 from the UK Twin Study. Fasting serum samples were analyzed using non-targeted mass spectrometry analysis. Metabolon, Inc. was employed for standardized processes of identification and relative qualification (<https://www.metabolon.com/>) [12]. A total of 486 metabolites were analyzed, comprising 177 unknown metabolites and 309 known metabolites, which were further classified into eight biochemical classes: peptide, nucleotide, amino acid, energy, cofactors and vitamins, lipid, carbohydrate, and xenobiotics. On the other hand, the GWAS summary statistics for total and sex-stratified VAT utilized MRI scans, including 20,038 women and 19,038 men, were obtained from the UK Biobank (<https://cvd.hugeamp.org/>) [13]. These scans were annotated using deep learning techniques. The GWAS was conducted using the UK Biobank imputed genotypes version 3, excluding SNPs with a minor allele frequency less than 1% and imputation quality below 0.9. The GWAS analysis was performed using BOLT-LMM v2.3.4. To account for potential confounding factors, the measures were adjusted for BMI, height, age at the time of MRI, age squared, sex, the first 10 principal components of genetic ancestry, genotyping array, and MRI center. Significant sex differences were observed, with men having a higher mean VAT of 5.0 L compared to 2.6 L in women. Lastly, we have also gathered the GWAS summary statistics on BMI and HbA1c from European populations, sourced respectively from the Genetic Investigation of ANthropometric Traits (GIANT) Consortium (https://portals.broadinstitute.org/collaboration/giant/index.php/GIANT_consortium) and the Meta-Analyses of Glucose and Insulin-related Traits Consortium (MAGIC) (<https://magicinvestigators.org/>) [14, 15].

Selection criteria for instrumental variables

The genetic variants were extracted using association thresholds of $P < 1e-5$, which are commonly used in MR analysis when there are limited SNPs available for the exposure variable. Linkage disequilibrium (LD) analysis ($r^2 < 0.1$, clumping distance = 500 kb) was conducted to minimize the impact of SNP associations based on the European 1000 Genomes Project Phase 3 reference panel. We further harmonized SNPs for exposure and outcome, and palindromic effects and allelic inconsistent SNPs were removed. IVs that were strongly associated with outcomes ($P < 5e-8$) were also removed. To assess the suitability of the identified IVs for representing metabolite levels, we calculated F-statistics and excluded IVs with F-statistics < 10 . The F-statistics formula employed was $R^2 (N-K-1) / [K (1-R^2)]$, where R^2 represents the explained variance of the exposure by the IVs,

N represents the effective sample size, and K indicates the count of variants included in the IV model. Additionally, the PhenoScanner online platform was employed to identify and remove SNPs associated with potential confounding factors (age, BMI, height, diabetes, hypertension, and nonalcoholic fatty liver disease) (<http://www.phenoscanter.medschl.cam.ac.uk>) [16]. Finally, to evaluate the statistical power, we utilized the online tool available at <https://shiny.cns.genomics.com/mRnd/> [17].

MR analysis

The MR analysis was conducted using the R software, primarily utilizing packages such as “TwoSampleMR”, “MR-PRESSO”, and “MendelianRandomization”. To determine the causal effects of blood metabolites and VAT, five commonly used MR methods were employed: inverse variance weighted (IVW), weighted median, simple mode, weighted mode, and MR-Egger regression analysis. The fixed-effects IVW method, which combines Wald ratios for each SNP to calculate a pooled estimate, was used as the primary method. The other methods were used as additional measures to support the findings. False Discovery Rate (FDR) correction was employed to control for false positives in multiple testing. A statistically significant correlation was defined as having an estimated causal effect with $FDR < 5\%$. Blood metabolites with an P value < 0.05 , but not reaching the FDR threshold, were deemed to potentially have a causal effect. Additionally, the causal relationship between identified metabolites with sex-stratified VAT was also explored. Heterogeneity between IVs was quantified using Cochran’s Q test. If $P < 0.05$, indicating the presence of heterogeneity, the random-effects IVW was used instead of the fixed-effects IVW [18]. The intercept of MR Egger regression was examined to assess the presence of underlying pleiotropy, with a $P < 0.05$ suggesting the directional pleiotropy. The MR-PRESSO test was employed to identify and quantify potential pleiotropic effects, detect outliers that could impact the study outcomes, and assess improvements after their removal. Leave-one-out analysis was also conducted to evaluate the influence of potentially significant IVs. Lastly, to unveil the potential vertical pleiotropic pathways of the identified blood metabolites, we conducted multivariable MR (MVMR) analyses, including MVMR-IVW, MVMR-Egger, and MVMR-Median, to estimate the direct causal effects of these blood metabolites on VAT after adjusting for BMI and HbA1c. The parameter settings were consistent with those of univariable MR analysis.

Metabolic pathway enrichment analysis

Based on the identified blood metabolites with P value < 0.05 , MetaboAnalyst 5.0 (<https://www.metaboanalyst.ca/>), an intuitive online tool specifically designed

for streamlined metabolomics data analysis, was used to conduct metabolic pathway analysis [19, 20].

Evaluation of genetic correlation and directionality

Genetic correlation between the exposure and outcome in MR analysis may lead to a violation of cause-effects [21, 22]. Despite excluding SNPs linked to VAT when choosing IVs, irrelevant SNPs might still influence VAT presence. LDSC is a statistical method used to analyze genetic correlation. By leveraging linkage disequilibrium information in the genome, it can assess the genetic correlation between two traits, such as disease and gene expression [21, 23]. To confirm that the causal effects are not muddled by shared genetics, LDSC was used to assess the genetic correlation between metabolites and VAT [24]. Furthermore, the MR Steiger test was conducted to address the potential bias arising from reverse causality [25].

Reverse MR analysis

Based on the same criteria, we conducted a reverse MR analysis using VAT-associated SNPs as IVs to investigate if VAT accumulation causally affected the blood metabolites identified above.

Results

Selection of instrumental variables

After conducting a rigorous quality control process, we have identified 10,635 SNPs linked to 486 blood metabolites as IVs, ranging from 3 to 505, to explore the causal relationship between metabolites and VAT. F-statistics for each SNP were all over 10, suggesting no weak IVs were employed. Comprehensive information for each SNP was presented in Supplementary 1.

Results of MR analysis between blood metabolites and VAT

The comprehensive analysis results of 486 blood metabolites and VAT was described in Supplementary 2. The IVW analysis identified a total of forty-five metabolites associated with VAT accumulation ($P < 0.05$). Among them, six metabolites remained chemically unknown. The remaining metabolites were assigned to eight amino acids, three carbohydrates, twenty-one lipids, two peptides, and five xenobiotics (Fig. 1). Based on the pathway analysis conducted using MetaboAnalyst for the identified blood metabolites, the results indicated that the top twenty-five ranked metabolic pathways are caffeine metabolism, glutathione metabolism, alpha linolenic acid and linoleic acid metabolism, and so on (Fig. 2).

Further after FDR correction, the IVW MR estimation identified a significance positive correlation between arachidonate (20:4n6) and the accumulation of VAT ($\beta = 0.35$, 95%CI:0.18–0.52, $P < 0.001$; $FDR = 0.025$). We also identified four metabolites with obvious potential

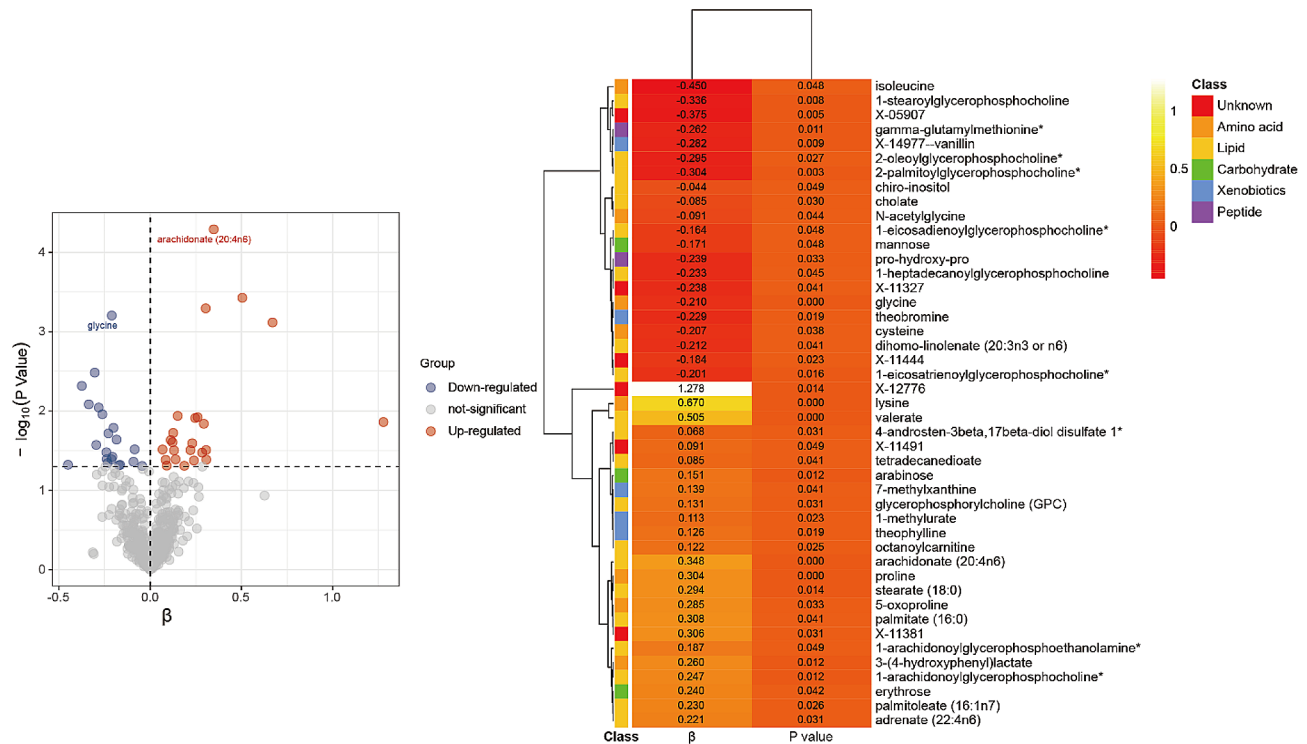


Fig. 1 Causal relationships between forty-five metabolites and VAT

causal relationship ($FDR < 0.10$). Higher levels of lysine ($\beta = 0.67$, 95%CI: 0.28–1.06, $P < 0.001$; $FDR = 0.074$), proline ($\beta = 0.30$, 95%CI: 0.13–0.48, $P < 0.001$; $FDR = 0.082$), and valerate ($\beta = 0.50$, 95%CI: 0.23–0.78, $P < 0.001$, $FDR = 0.091$) might also result in the accumulation of VAT. On the other hand, the presence of glycine ($\beta = -0.21$, 95%CI: -0.33–0.09), $P < 0.001$, $FDR = 0.076$) suggested a protective effect against the accumulation of VAT (Fig. 3). MR Egger, weighted median, simple mode, and weighted mode also demonstrated a trend of causal relationships. Supplementary 3 displayed scatterplots demonstrating the causal relationship between metabolites and VAT accumulation. Cochran's Q test suggested there is no indication of heterogeneity among SNPs associated with blood metabolites in predicting VAT accumulation ($P > 0.05$) (Table 1). MR-Egger regression intercept also showed no risk discrepancy due to unbalanced pleiotropy related to VAT accumulation ($P > 0.05$) (Table 1). The MR-PRESSO test further confirmed the reliability of our results ($P > 0.05$) (Table 1). Lastly, leave-one-out analysis, as shown in Supplementary 3, did not find any influential SNPs affecting the overall effect estimate.

Results of MR analysis between blood metabolites and sex-stratified VAT

We further investigated the relationship between above five blood metabolites and VAT stratified by sex (Fig. 4). Our analysis showed consistent trends between different

sexes for most blood metabolites. However, the association between arachidonate (20:4n6), lysine, proline, valerate and VAT accumulation was statistically different in females ($P < 0.05$), while the associations between valerate, glycine and VAT accumulation were statistically different in males ($P < 0.05$).

MVMR analysis

We further employed a multivariable MR analysis to estimate the direct effects of these five metabolites on the accumulation of VAT (Fig. 5). Based on the MVMR_IVW results, the effect of genetically predicted arachidonate (20:4n6) ($\beta = 0.28$, 95%CI: 0.05–0.52, $P = 0.017$) and proline ($\beta = 0.40$, 95%CI: 0.15–0.66, $P = 0.002$) on VAT remained after accounting for BMI and HbA1c. The causal inference was further supported by consistent direction and magnitude from the results of MVMR_Egger and MVMR_Median model.

Results of MR analysis at genome-wide significance threshold ($5e-8$)

Upon further refinement based on genome-wide significance thresholds ($5e-8$) with LD analysis ($r^2 < 0.001$, clumping distance = 10,000 kb), we selected IVs for five blood metabolites (Fig. 6). arachidonate (20:4n6), glycine, and lysine each had one SNPs as IVs; proline had three SNPs as potential IVs. However, for valerate, no suitable IVs were found. Further IVW analysis indicated that the

Metabolic pathways (top 25)

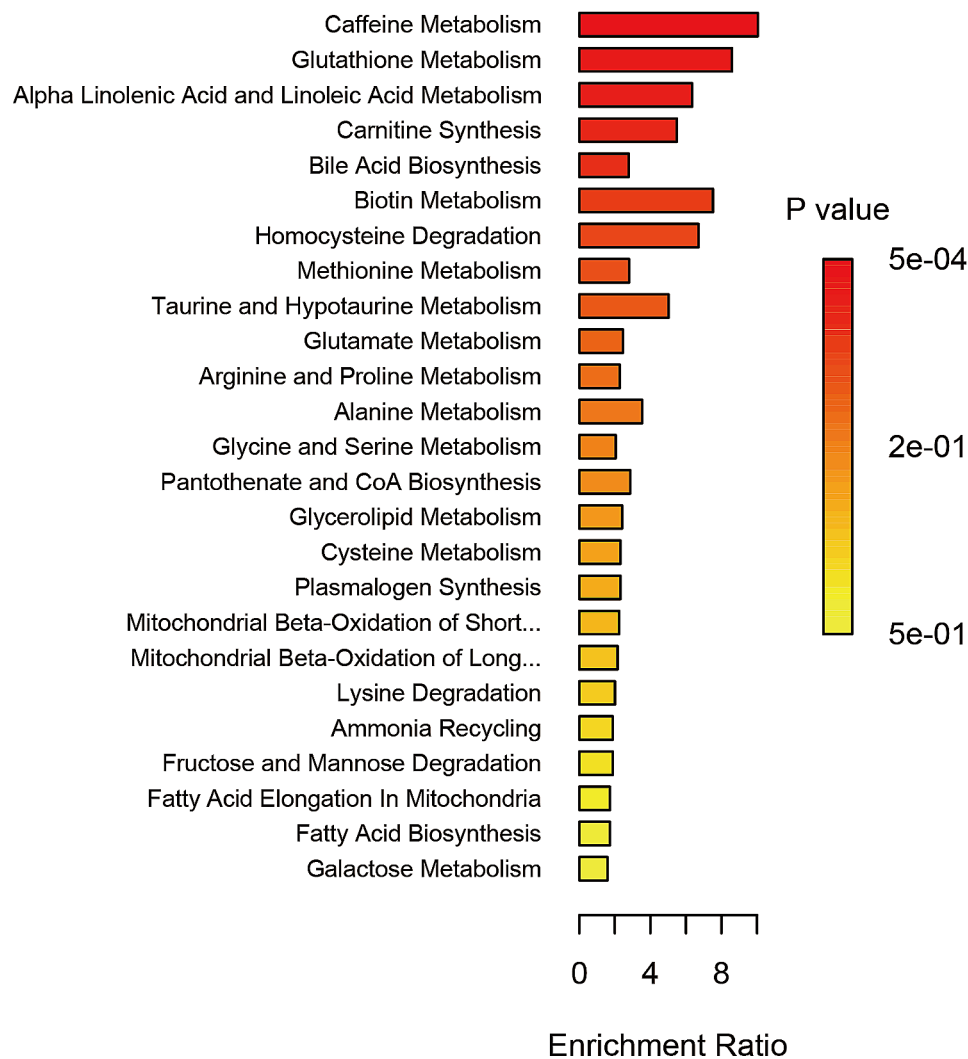


Fig. 2 Results of pathway enrichment analysis of forty-five metabolites

increased levels of arachidonate (20:4n6) ($\beta=0.40$, 95%CI: 0.10–0.71, $P=0.010$), lysine ($\beta=1.96$, 95%CI: 0.87–3.05, $P<0.001$), and proline ($\beta=0.40$, 95%CI: 0.09–0.72, $P=0.013$) remain as the risk factors for the augmented accumulation of VAT.

Evaluation of genetic correlation and directionality

To determine if the association between metabolites and VAT is due to shared genetic structure, we conducted LDSC analysis (Supplementary 3). Based on the total VAT, the results indicate no significant genetic correlation between arachidonate (20:4n6), glycine, lysine, valerate, and VAT accumulation. However, there may be a genetic correlation between VAT and proline ($R_g=0.190$, $P=0.011$). It is important to acknowledge that the limited

sample size of the metabolites may have compromised the statistical power of our analysis. Steiger test indicated no reverse causation bias in the identified causal relationships ($P<0.05$) (Supplementary 3). Finally, reverse MR analysis did not find evidence of the causal relationship between VAT accumulation and the five blood metabolites mentioned above (Supplementary 2). For other potential causal metabolites, reverse MR analyses results indicated that suggest that there is a bidirectional causal relationship between the accumulation of VAT and arabinose, N-acetylglycine, 1-stearoylglycerophosphocholine, 2-palmitoylglycerophosphocholine*, chiro-inositol.

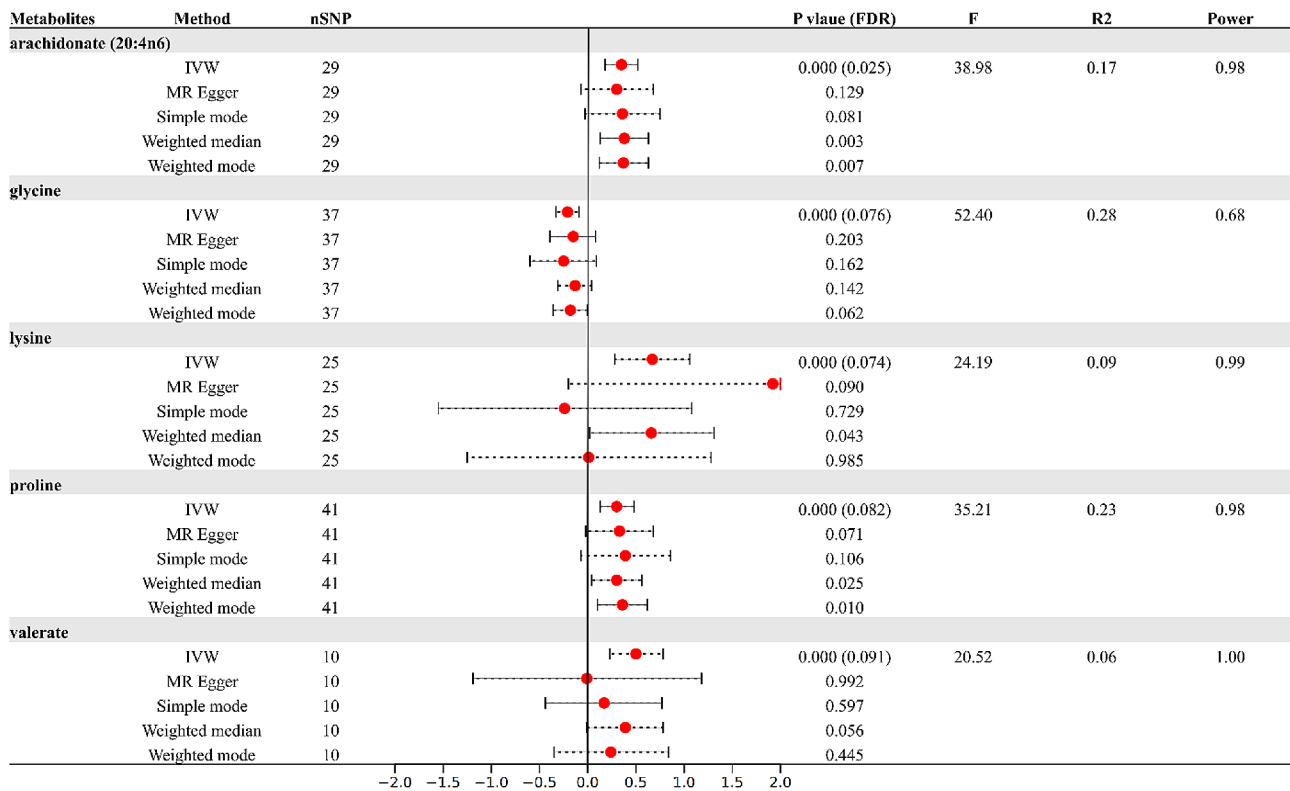


Fig. 3 Causal relationships between five metabolites and VAT

Table 1 The results of Cochran’s Q, MR-Egger intercept, and MR-PRESSO

Metabolites	Group	Q _{IVW}	P	Egger intercept	P	MR-PRESSO global test
arachidonate (20:4n6)	Total	38.830	0.084	0.001	0.768	0.134
	Female	43.163	0.034	-0.004	0.470	0.051
	Male	32.222	0.266	0.006	0.242	0.324
glycine	Total	40.928	0.263	-0.002	0.571	0.290
	Female	29.018	0.789	0.001	0.804	0.829
	Male	44.190	0.164	-0.003	0.421	0.166
lysine	Total	29.104	0.216	-0.012	0.251	0.217
	Female	27.727	0.272	-0.011	0.453	0.266
	Male	21.492	0.610	-0.013	0.354	0.612
proline	Total	51.642	0.103	-0.001	0.847	0.136
	Female	57.891	0.033	0.000	0.901	0.055
	Male	36.315	0.637	-0.001	0.666	0.677
valerate	Total	15.489	0.078	0.011	0.399	0.114
	Female	15.366	0.081	0.013	0.504	0.108
	Male	7.494	0.586	0.007	0.617	0.647

Discussion

To our knowledge, this is the first MR study to assess the causal role of human blood metabolites in visceral obesity.

Obesity is a multifactorial disease with significant heterogeneity. It manifests in various phenotypes, which can be either metabolically unhealthy or healthy [26]. Metabolomics, a high-throughput, and unbiased profiling technique, enables the simultaneous quantification

of a wide range of small-molecule metabolites within biological systems [27]. By analyzing the metabolome, which represents the end-products of cellular processes, metabolomics offers a unique opportunity to uncover the intricate metabolic alterations associated with obesity [28]. We have identified that the metabolic pathways primarily enriched in these metabolites include caffeine metabolism. Current research consistently suggests a strong correlation between caffeine metabolism

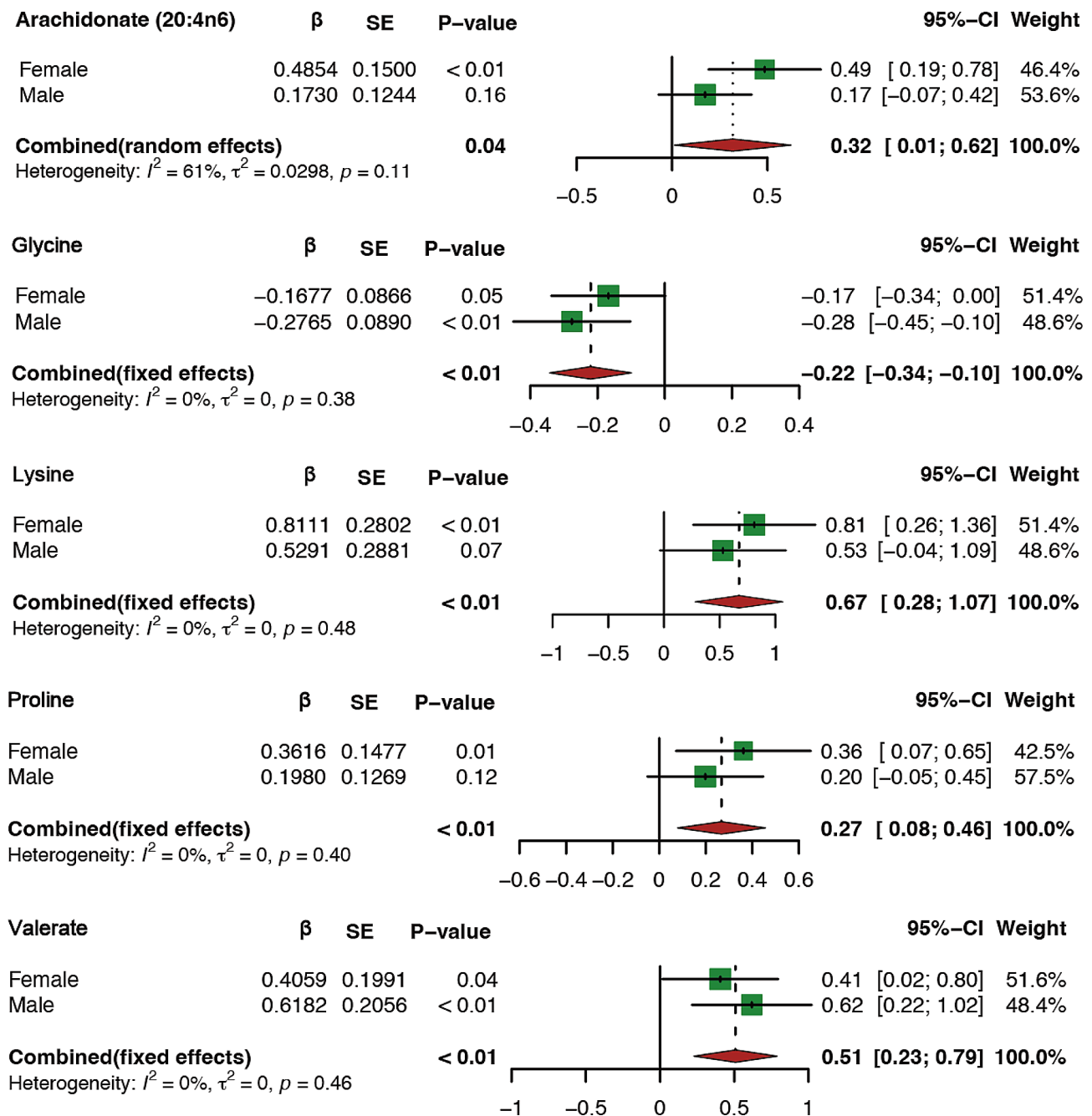


Fig. 4 Causal relationships between five metabolites and sex-stratified VAT

and obesity, including anti-inflammatory and antioxidant effects that promote fat oxidation, increase energy metabolism, and suppress appetite [29–31]. Previous non-targeted metabolomics study focusing on the obese people and animals also revealed that caffeine and caffeine-related metabolism pathways were the most prominent metabolic pathways [32, 33]. Additionally, previous studies have suggested that some of these blood metabolites, such as arachidonate (20:4n6) [34], glycine [35], palmitoleate [36], theophylline [37], and so on, might be

involved in the pathogenesis of obesity and serve as biomarkers for obesity. However, most of these studies have focused on general obesity measures such as BMI, while the distribution of body adipose tissue plays a crucial role in obesity and its associated health risks. By leveraging previous GWAS studies, we can more accurately identify the harmful obesity phenotype, specifically visceral obesity. Furthermore, based on the advantages MR studies, we have further elucidated the pathogenic and protective effects of these blood metabolites on the

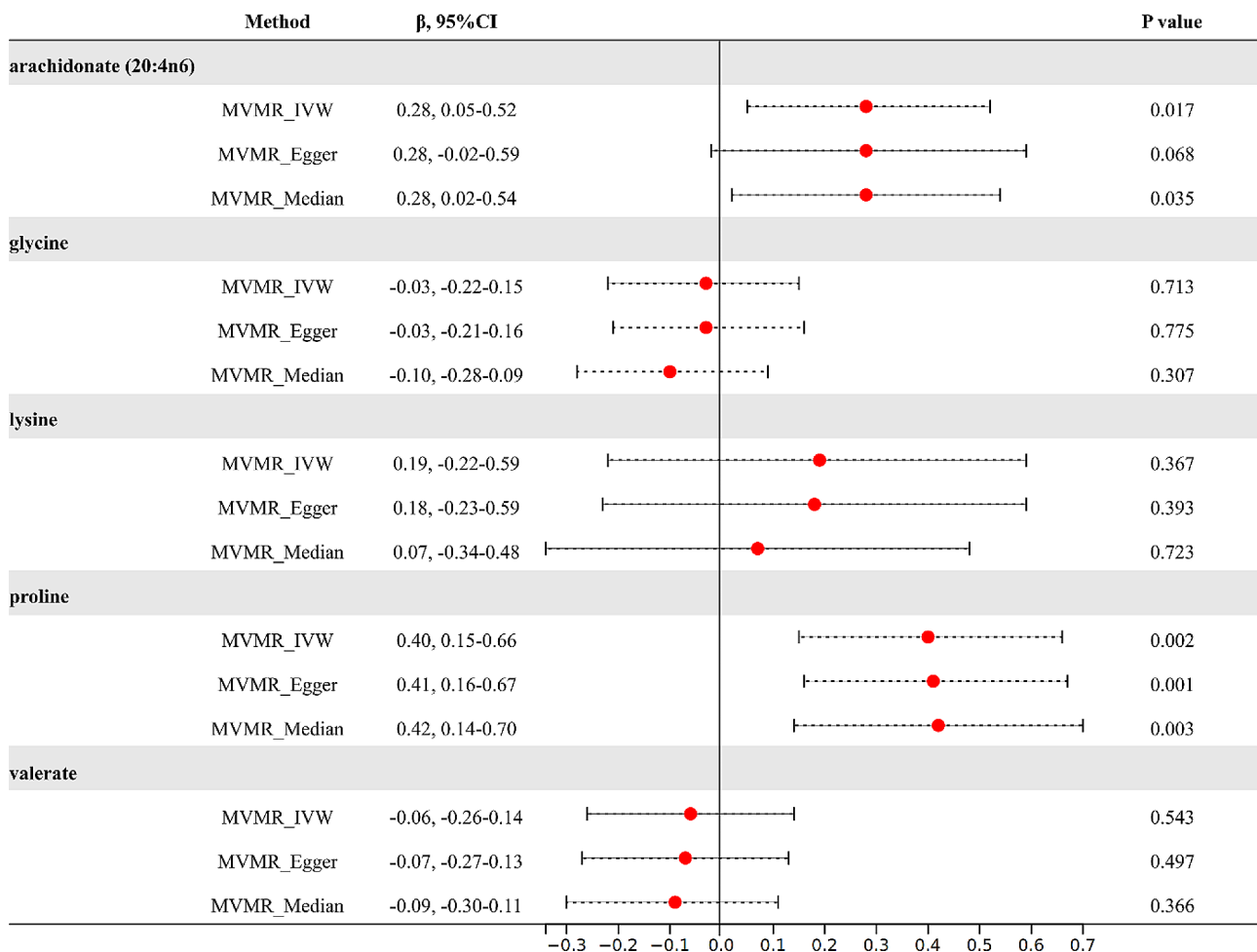


Fig. 5 Results of MVMR analysis

accumulation of VAT. This additional information provides a more comprehensive understanding of the role of these blood metabolites in visceral obesity and related health outcomes.

After multiple testing correction, arachidonate (20:4n6) has been implicated in VAT accumulation. Arachidonate is a crucial essential fatty acid in the human body, and it is the most abundant and widely distributed polyunsaturated fatty acid. The metabolic products of arachidonate include a series of prostaglandins and leukotrienes, which are highly active inflammatory mediators. Studies have shown that it has a pro-inflammatory effect in the inflammatory microenvironment of 3T3-L1 adipocytes induced by lipopolysaccharides [38]. Additionally, four other blood metabolites (glycine, lysine, proline, and valerate) were identified as having obvious potential causal relationship ($FDR < 0.10$). Glycine, the amino acid with the lowest molecular weight, shows lower circulating levels in metabolic disorders related to obesity, type 2 diabetes (T2DM), and non-alcoholic fatty liver disease (NAFLD), and increasing glycine levels can inhibit these

disorders in clinical application [35]. Lysine acetylation plays a crucial role in both immune and metabolic pathways, regulating the balance of energy storage and expenditure. Current evidence suggests that lysine acetylation can modulate innate immune and metabolic pathways related to obesity and metabolic diseases [39]. Proline, playing a role in the regulation of food intake and body fat accumulation suggests its potential as a target for interventions in managing visceral obesity [40]. However, there is limited research on the relationship between valerate and VAT accumulation and obesity. Further exploration is needed in the future to better understand the potential impact of valerate on visceral obesity.

There are several limitations to this study that may impact the interpretation of the results. Firstly, the SNP used did not meet the GWAS significance threshold ($5e-8$). Although we relaxed the selection criteria for IVs, it was considered an acceptable threshold for blood metabolites and has been adopted in other articles [23, 41]. Secondly, most participants in our study were of European descent. While this helped to minimize population

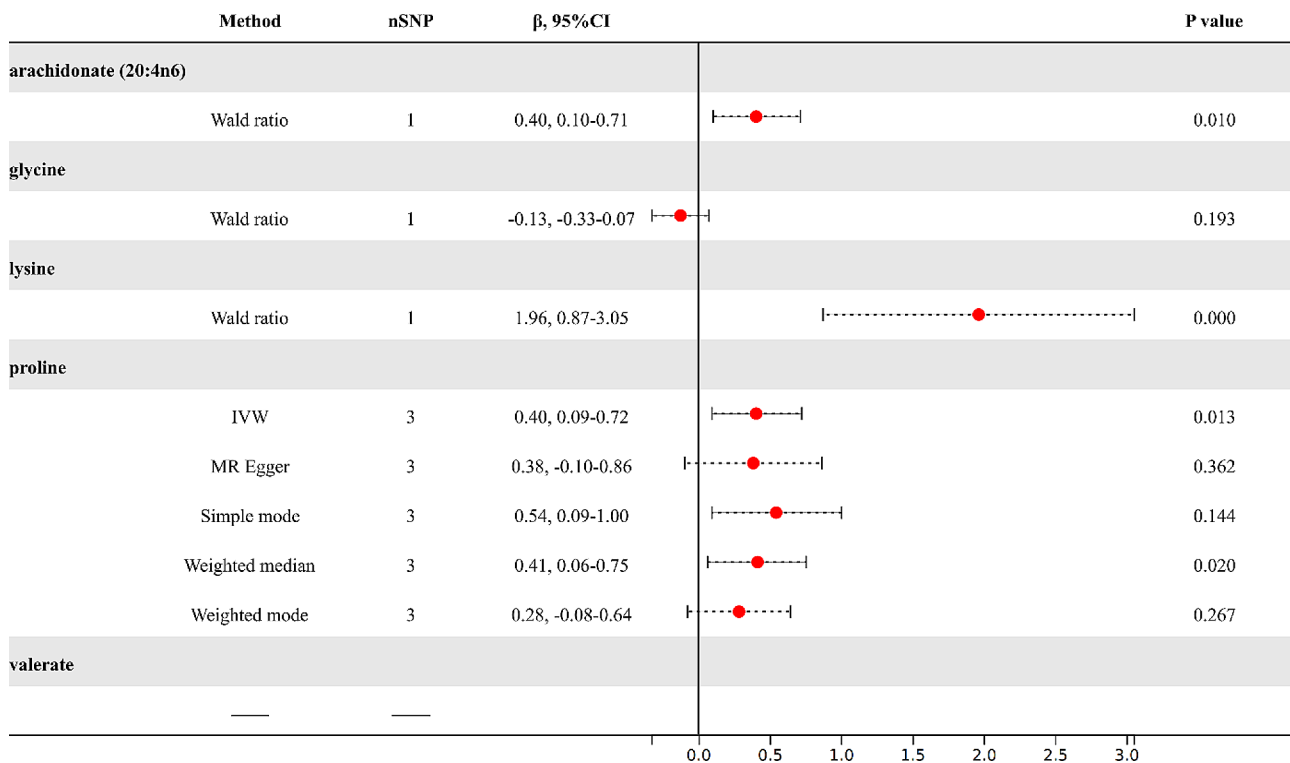


Fig. 6 Results of MR analysis at genome-wide significance threshold ($5e-8$)

heterogeneity, it is important to validate the MR results in other populations to ensure their generalizability. Thirdly, VAT, due to its metabolic activity, could induce changes in the metabolome. We conducted a reverse MR analysis only on metabolites with potential causal relationships, and future research should delve deeper into the impact of VAT on human metabolism from this perspective. Furthermore, conducting further MR analyses, such as MR-RAPS, and reinforcing the reliability of results through replication validation based on external populations, are also necessary. Lastly, while the MR approach is excellent for causal inference, it is crucial to validate the findings from this study in well-powered randomized controlled trials.

Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s12944-024-02035-x>.

Supplementary Material 1: The detailed characteristics of IVs

Supplementary Material 2: MR analysis results

Supplementary Material 3: The results of scatterplots, leave-one-out, LDSC, and Steiger test

Acknowledgements

We want to acknowledge all participants of this study and the technical support provided by the Jiangsu University.

Author contributions

Z.W. and Q.Y. wrote the main manuscript text and prepared figures and tables. All authors reviewed the manuscript.

Funding

This study was funded by the Science and Technology Project of Changzhou Health Commission (WZ202226).

Data availability

No datasets were generated or analysed during the current study.

Declarations

Ethics approval

Not applicable.

Informed consent

Not applicable.

Competing interests

The authors declare no competing interests.

Received: 27 November 2023 / Accepted: 30 January 2024

Published online: 07 February 2024

References

- Zhang X, Ha S, Lau HC, Yu J. Excess body weight: novel insights into its roles in obesity comorbidities. *Semin Cancer Biol.* 2023;9216–27. <https://doi.org/10.1016/j.semcancer.2023.03.008>.
- Ibrahim MM. Subcutaneous and visceral adipose tissue: structural and functional differences. *Obes Rev.* 2010;11(1):11–8. <https://doi.org/10.1111/j.1467-789X.2009.00623.x>.
- Neeland IJ, Ross R, Després JP, Matsuzawa Y, Yamashita S, Shai I, Seidell J, Magni P, Santos RD, Arsenault B, Cuevas A, Hu FB, Griffin B, Zambon A, Barter P, Fruchart JC, Eckel RH. Visceral and ectopic fat, atherosclerosis, and

- cardiometabolic disease: a position statement. *Lancet Diabetes Endocrinol.* 2019;7(9):715–25. [https://doi.org/10.1016/s2213-8587\(19\)30084-1](https://doi.org/10.1016/s2213-8587(19)30084-1).
4. Silveira EA, Kliemann N, Noll M, Sarrafzadegan N, de Oliveira C. Visceral obesity and incident cancer and cardiovascular disease: an integrative review of the epidemiological evidence. *Obes Rev.* 2021;22(1):e13088. <https://doi.org/10.1111/obr.13088>.
 5. Doyle SL, Donohoe CL, Lysaght J, Reynolds JV. Visceral obesity, metabolic syndrome, insulin resistance and cancer. *Proc Nutr Soc.* 2012;71(1):181–9. <https://doi.org/10.1017/s002966511100320x>.
 6. Johnson CH, Ivanisevic J, Siuzdak G. Metabolomics: beyond biomarkers and towards mechanisms. *Nat Rev Mol Cell Biol.* 2016;17(7):451–9. <https://doi.org/10.1038/nrm.2016.25>.
 7. Yang Q, Vijayakumar A, Kahn BB. Metabolites as regulators of insulin sensitivity and metabolism. *Nat Rev Mol Cell Biol.* 2018;19(10):654–72. <https://doi.org/10.1038/s41580-018-0044-8>.
 8. Sekula P, Del Greco MF, Pattaro C, Köttgen A. Mendelian randomization as an Approach to assess causality using Observational Data. *J Am Soc Nephrol.* 2016;27(11):3253–65. <https://doi.org/10.1681/asn.2016010098>.
 9. Skrivankova VV, Richmond RC, Woolf BAR, Yarmolinsky J, Davies NM, Swanson SA, VanderWeele TJ, Higgins JPT, Timpson NJ, Dimou N, Langenberg C, Golub RM, Loder EW, Gallo V, Tybjaerg-Hansen A, Davey Smith G, Egger M, Richards JB. Strengthening the reporting of Observational studies in Epidemiology using mendelian randomization: the STROBE-MR Statement. *JAMA.* 2021;326(16):1614–21. <https://doi.org/10.1001/jama.2021.18236>.
 10. Buniello A, MacArthur JAL, Cerezo M, Harris LW, Hayhurst J, Malangone C, McMahon A, Morales J, Mountjoy E, Solis E, Suveges D, Vrousou O, Whetzel PL, Amode R, Guillen JA, Riat HS, Trevanion SJ, Hall P, Junkins H, Flicek P, Burdett T, Hindorf LA, Cunningham F, Parkinson H. The NHGRI-EBI GWAS catalog of published genome-wide association studies, targeted arrays and summary statistics 2019. *Nucleic Acids Res.* 2019;47(D1):D1005–d1012. <https://doi.org/10.1093/nar/gky1120>.
 11. Shin SY, Fauman EB, Petersen AK, Krumsiek J, Santos R, Huang J, Arnold M, Erte I, Forgetta V, Yang TP, Walter K, Menni C, Chen L, Vasquez L, Valdes AM, Hyde CL, Wang V, Ziemek D, Roberts P, Xi L, Grundberg E, Waldenberger M, Richards JB, Mohny RP, Milburn MV, John SL, Trimmer J, Theis FJ, Overington JP, Suhre K, Brosnan MJ, Gieger C, Kastenmüller G, Spector TD, Soranzo N. An atlas of genetic influences on human blood metabolites. *Nat Genet.* 2014;46(6):543–50. <https://doi.org/10.1038/ng.2982>.
 12. Ryals J, Lawton K, Stevens D, Milburn M. Metabolon Inc Pharmacogenomics. 2007;8(7):863–6. <https://doi.org/10.2217/14622416.8.7.863>.
 13. Agrawal S, Wang M, Klarqvist MDR, Smith K, Shin J, Dashti H, Diamant N, Choi SH, Jurgens SJ, Ellinor PT, Philippakis A, Claussnitzer M, Ng K, Udler MS, Batra P, Khera AV. Inherited basis of visceral, abdominal subcutaneous and gluteofemoral fat depots. *Nat Commun.* 2022;13(1):3771. <https://doi.org/10.1038/s41467-022-30931-2>.
 14. Wheeler E, Leong A, Liu CT, Hivert MF, Strawbridge RJ, Podmore C, Li M, Yao J, Sim X, Hong J, Chu AY, Zhang W, Wang X, Chen P, Maruthur NM, Porneala BC, Sharp SJ, Jia Y, Kabagambe EK, Chang LC, Chen WM, Elks CE, Evans DS, Fan Q, Giulianini F, Go MJ, Hottenga JJ, Hu Y, Jackson AU, Kanoni S, Kim YJ, Kleber ME, Ladenvall C, Leccoer C, Lim SH, Lu Y, Mahajan A, Marzi C, Nalls MA, Navarro P, Nolte IM, Rose LM, Rybin DV, Sanna S, Shi Y, Stram DO, Takeuchi F, Tan SP, van der Most PJ, Van Vliet-Ostapchouk JV, Wong A, Yengo L, Zhao W, Goel A, Martinez Larrad MT, Radke D, Salo P, Tanaka T, van Iperen EPA, Abecasis G, Afari S, Alizadeh BZ, Bertoni AG, Bonnefond A, Böttcher Y, Bottinger EP, Campbell H, Carlson OD, Chen CH, Cho YS, Garvey WT, Gieger C, Goodarzi MO, Grallert H, Hamsten A, Hartman CA, Herder C, Hsiung CA, Huang J, Igase M, Isona M, Katsuya T, Khor CC, Kiess W, Kohara K, Kovacs P, Lee J, Lee WJ, Lehne B, Li H, Liu J, Lobbens S, Luan J, Lyssenko V, Meitinger T, Miki T, Miljkovic I, Moon S, Mulas A, Müller G, Müller-Nurasyid M, Nagaraja R, Nauck M, Pankow JS, Polasek O, Prokopenko I, Ramos PS, Rasmussen-Torvik L, Rathmann W, Rich SS, Robertson NR, Roden M, Rousset R, Rudan I, Scott RA, Scott WR, Sennblad B, Siscovick DS, Strauch K, Sun L, Swertz M, Tajuddin SM, Taylor KD, Teo YY, Tham YC, Tönjes A, Wareham NJ, Willemssen G, Wilsaard T, Hingorani AD, Egan J, Ferrucci L, Hovingh GK, Jula A, Kivimäki M, Kumari M, Njølstad I, Palmer CNA, Soranzo Rios M, Stumvoll M, Watkins H, Aung T, Blüher M, Boehnke M, Boomsma DI, Bornstein SR, Chambers JC, Chasman DJ, Chen YI, Chen YT, Cheng CY, Cucca F, de Geus EJC, Deloukas P, Evans MK, Fornage M, Friedlander Y, Froguel P, Groop L, Gross MD, Harris TB, Hayward C, Heng CK, Ingelsson E, Kato N, Kim BJ, Koh WP, Kooper JS, Körner A, Kuh D, Kuusisto J, Laakso M, Lin X, Liu Y, Loos RFJ, Magnusson PKE, März W, McCarthy MI, Oldehinkel AJ, Ong KK, Pedersen NL, Pereira MA, Peters A, Ridker PM, Sabanayagam C, Sale M, Saleheen D, Saltevo J, Schwarz PE, Sheu WHH, Snieder H, Spector TD, Tabara Y, Tuomilehto J, van Dam RM, Wilson JG, Wilson JF, Wolfenbuttel BHR, Wong TY, Wu JY, Yuan JM, Zonderman AB, Soranzo N, Guo X, Roberts DJ, Florez JC, Sladek R, Dupuis J, Morris AP, Tai ES, Selvin E, Rotter JJ, Langenberg C, Barroso I, Meigs JB (2017) Impact of common genetic determinants of Hemoglobin A1c on type 2 diabetes risk and diagnosis in ancestrally diverse populations: A transethnic genome-wide meta-analysis. *PLoS Med* 14(9): e1002383. <https://doi.org/10.1371/journal.pmed.1002383>.
 15. Locke AE, Kahali B, Berndt SI, Justice AE, Pers TH, Day FR, Powell C, Vedantam S, Buchkovich ML, Yang J, Croteau-Chonka DC, Esko T, Fall T, Ferreira T, Gustafsson S, Kutalik Z, Luan J, Mägi R, Randall JC, Winkler TW, Wood AR, Workalemahu T, Faul JD, Smith JA, Zhao JH, Zhao W, Chen J, Fehrmann R, Hedman Å K, Karjalainen J, Schmidt EM, Absher D, Amin N, Anderson D, Beekman M, Bolton JL, Bragg-Gresham JL, Buyske S, Demirkan A, Deng G, Ehret GB, Feenstra B, Feitosa MF, Fischer K, Goel A, Gong J, Jackson AU, Kanoni S, Kleber ME, Kristiansson K, Lim U, Lotay V, Mangino M, Leach IM, Medina-Gomez C, Medland SE, Nalls MA, Palmer CD, Pasko D, Pechlivanis S, Peters MJ, Prokopenko I, Shungin D, Stancáková A, Strawbridge RJ, Sung YJ, Tanaka T, Teumer A, Trompet S, van der Laan SW, van Setten J, Van Vliet-Ostapchouk JV, Wang Z, Yengo L, Zhang W, Isaacs A, Albrecht E, Ärnlöv J, Ascott GM, Attwood AP, Bandinelli S, Barrett A, Bas IN, Bellis C, Bennett AJ, Berne C, Blagieva R, Blüher M, Böhringer S, Bonnycastle LL, Böttcher Y, Boyd HA, Bruinenberg M, Caspersen IH, Chen YI, Clarke R, Daw EW, de Craen AJM, Delgado G, Dimitriou M, Doney ASF, Eklund N, Estrada K, Eury E, Folkersen L, Fraser RM, Garcia ME, Geller F, Giedraitis V, Gigante B, Go AS, Golay A, Goodall AH, Gordon SD, Gorski M, Grabe HJ, Grallert H, Grammer TB, Gräßler J, Grönberg H, Groves CJ, Gusto G, Haessler J, Hall P, Haller T, Hallmans G, Hartman CA, Hassinen M, Hayward C, Heard-Costa NL, Helmer Q, Hengstenberg C, Holmen O, Hottenga JJ, James AL, Jeff JM, Johansson Å, Jolley J, Juliusdottir T, Kinnunen L, Koenig W, Koskenvuo M, Kratzer W, Laitinen J, Lamina C, Leander K, Lee NR, Lichtner P, Lind L, Lindström J, Lo KS, Lobbens S, Lorbeer R, Lu Y, Mach F, Magnusson PKE, Mahajan A, McArdle WL, McLachlan S, Menni C, Merger S, Mihailov E, Milani L, Moayeri A, Monda KL, Morken MA, Mulas A, Müller G, Müller-Nurasyid M, Musk AW, Nagaraja R, Nöthen MM, Nolte IM, Pilz S, Rayner NW, Renstrom F, Rettig R, Ried JS, Ripke S, Robertson NR, Rose LM, Sanna S, Schragmahl J, Scholtens S, Schumacher FR, Scott WR, Seufferlein T, Shi J, Smith AV, Smolonska J, Stanton AV, Steinthorsdottir V, Stirrups K, Stringham HM, Sundström J, Swertz MA, Swift AJ, Svyänen AC, Tan ST, Tayo BO, Thorand B, Thorleifsson G, Tyrer JP, Uh HW, Vandenput L, Verhulst FC, Vermeulen SH, Verweij N, Vonk JM, Waite LL, Warren HR, Waterworth D, Weedon MN, Wilkens LR, Willenborg C, Wilsaard T, Wojczynski MK, Wong A, Wright AF, Zhang Q, Brennan EP, Choi M, Dastani Z, Drong AW, Eriksson P, Franco-Cereceda A, Gädin JR, Gharavi AG, Goddard ME, Handsaker RE, Huang J, Karpe F, Kathiresan S, Keildson S, Kiryluk K, Kubo M, Lee JY, Liang L, Lifton RP, Ma B, McCarroll SA, McCann AJ, Mian J, Moffatt MF, Montgomery GW, Murabito JM, Nicholson G, Nyholt DR, Okada Y, Perry JRB, Dorajoo R, Reinmaa E, Saleem RM, Sandholm N, Scott RA, Stolk L, Takahashi A, Tanaka T, van 't Hooft FM, Vinkhuyzen AAE, Westra HJ, Zheng W, Zondervan KT, Heath AC, Arveiler D, Bakker SLL, Beilby J, Bergman RN, Blangero J, Bovet P, Campbell H, Caulfield MJ, Cesana G, Chakravarti A, Chasman DJ, Chines PS, Collins FS, Crawford DC, Cupples LA, Cusi D, Danesh J, de Faire U, den Ruijter HM, Dominiczak AF, Erbel R, Erdmann J, Eriksson JG, Farrall M, Felix SB, Ferrannini E, Ferrières J, Ford I, Forouhi NG, Forrester T, Franco OH, Gansevoort RT, Gejman PV, Gieger C, Gottesman O, Gudnason V, Gyllenstein U, Hall AS, Harris TB, Hattersley AT, Hicks AA, Hindorf LA, Hingorani AD, Hofman A, Homuth G, Hovingh GK, Humphries SE, Hunt SC, Hyppönen E, Illig T, Jacobs KB, Jarvelin MR, Jöckel KH, Johansen B, Jousilahti P, Jukema JW, Jula AM, Kaprio J, Kastelein JJP, Keinanen-Kiukkaanniemi SM, Kiemenev LA, Knekt PK, Kooper JS, Kooperberg C, Kovacs P, Kraja AT, Kumari M, Kuusisto J, Lakka TA, Langenberg C, Marchand LL, Lehtimäki T, Lyssenko V, Männistö S, Miettinen M, Matisse TC, McKenzie CA, McKnight B, Moll FL, Morris AD, Morris AP, Murray JC, Nelis M, Ohlsson C, Oldehinkel AJ, Ong KK, Madden PAF, Pasterkamp G, Peden JF, Peters A, Postma DS, Pramstaller PP, Price JF, Qi L, Raitakari OT, Rankinen T, Rao DC, Rice TK, Ridker PM, Rioux JD, Ritchie MD, Rudan I, Saloma V, Samani NJ, Saramies J, Sarzynski MA, Schunkert H, Schwarz PEH, Sever P, Shuldiner AR, Sinisalo J, Stolk RP, Strauch K, Tönjes A, Trégouët DA, Tremblay A, Tremoli E, Virtamo J, Vohl MC, Völker U, Waeber G, Willemssen G, Wittmann JC, Zillikens MC, Adair LS, Amouyel P, Asselbergs FW, Assimes TL, Bochud M, Boehm BO, Boerwinkle E, Bornstein SR, Bottinger EP, Boucharde C, Cauchi S, Chambers JC, Chanock SJ, Cooper RS, de Bakker PIW, Dedoussis G, Ferrucci L, Franks PW, Froguel P, Groop LC, Haiman CA, Hamsten A, Hui J, Hunter DJ, Hveem K, Kaplan RC, Kivimäki M, Kuh D, Laakso M, Liu Y, Martin NG, März W, Melbye M, Metspalu A, Moebus S, Munroe PB, Njølstad I, Oostra BA, Palmer CNA, Pedersen NL, Perola M, Pérusse L, Peters U, Power C, Quertermous T, Rauramaa

- R, Rivadeneira F, Saaristo TE, Saleheen D, Sattar N, Schadt EE, Schlessinger D, Slagboom PE, Snieder H, Spector TD, Thorsteinsdottir U, Stumvoll M, Tuomilehto J, Uitterlinden AG, Uusitupa M, van der Harst P, Walker M, Wallaschofski H, Wareham NJ, Watkins H, Weir DR, Wichmann HE, Wilson JF, Zanen P, Borecki IB, Deloukas P, Fox CS, Heid IM, O'Connell JR, Strachan DP, Stefansson K, van Duijn CM, Abecasis GR, Franke L, Frayling TM, McCarthy MI, Visscher PM, Scherag A, Willer CJ, Boehnke M, Mohlke KL, Lindgren CM, Beckmann JS, Barroso I, North KE, Ingelsson E, Hirschhorn JN, Loos RJF, Speliotes EK (2015) Genetic studies of body mass index yield new insights for obesity biology. *Nature* 518(7538): 197–206. <https://doi.org/10.1038/nature14177>.
16. Kamat MA, Blackshaw JA, Young R, Surendran P, Burgess S, Danesh J, Butterworth AS, Staley JR. PhenoScanner V2: an expanded tool for searching human genotype-phenotype associations. *Bioinformatics*. 2019;35(22):4851–3. <https://doi.org/10.1093/bioinformatics/btz469>.
 17. Brion MJ, Shakhbuzov K, Visscher PM. Calculating statistical power in mendelian randomization studies. *Int J Epidemiol*. 2013;42(5):1497–501. <https://doi.org/10.1093/ije/dyt179>.
 18. Bowden J, Spiller W, Del Greco MF, Sheehan N, Thompson J, Minelli C, Davey Smith G. Improving the visualization, interpretation and analysis of two-sample summary data mendelian randomization via the Radial plot and radial regression. *Int J Epidemiol*. 2018;47(6):2100. <https://doi.org/10.1093/ije/dyy265>.
 19. Pang Z, Chong J, Zhou G, de Lima Morais DA, Chang L, Barrette M, Gauthier C, Jacques P, Li S, Xia J. MetaboAnalyst 5.0: narrowing the gap between raw spectra and functional insights. *Nucleic Acids Res*. 2021;49(W1):W388–w396. <https://doi.org/10.1093/nar/gkab382>.
 20. Kanehisa M, Furumichi M, Tanabe M, Sato Y, Morishima K. KEGG: new perspectives on genomes, pathways, diseases and drugs. *Nucleic Acids Res*. 2017;45(D1):D353–d361. <https://doi.org/10.1093/nar/gkw1092>.
 21. O'Connor LJ, Price AL. Distinguishing genetic correlation from causation across 52 diseases and complex traits. *Nat Genet*. 2018;50(12):1728–34. <https://doi.org/10.1038/s41588-018-0255-0>.
 22. Reay WR, Kiltschewskij DJ, Geaghan MP, Atkins JR, Carr VJ, Green MJ, Cairns MJ. Genetic estimates of correlation and causality between blood-based biomarkers and psychiatric disorders. *Sci Adv*. 2022;8(14):eabj8969. <https://doi.org/10.1126/sciadv.abj8969>.
 23. Cai J, Li X, Wu S, Tian Y, Zhang Y, Wei Z, Jin Z, Li X, Chen X, Chen WX. Assessing the causal association between human blood metabolites and the risk of epilepsy. *J Transl Med*. 2022;20(1):437. <https://doi.org/10.1186/s12967-022-03648-5>.
 24. Bulik-Sullivan BK, Loh PR, Finucane HK, Ripke S, Yang J, Patterson N, Daly MJ, Price AL, Neale BM. LD score regression distinguishes confounding from polygenicity in genome-wide association studies. *Nat Genet*. 2015;47(3):291–5. <https://doi.org/10.1038/ng.3211>.
 25. Xiao G, He Q, Liu L, Zhang T, Zhou M, Li X, Chen Y, Chen Y, Qin C. Causality of genetically determined metabolites on anxiety disorders: a two-sample mendelian randomization study. *J Transl Med*. 2022;20(1):475. <https://doi.org/10.1186/s12967-022-03691-2>.
 26. Milhem F, Komarnytsky S. Progression to obesity: variations in patterns of metabolic fluxes, Fat Accumulation, and gastrointestinal responses. *Metabolites*. 2023;13(9). <https://doi.org/10.3390/metabo13091016>.
 27. Wishart DS. Metabolomics for investigating physiological and pathophysiological processes. *Physiol Rev*. 2019;99(4):1819–75. <https://doi.org/10.1152/physrev.00035.2018>.
 28. Newgard CB. Metabolomics and metabolic diseases: where do we stand? *Cell Metab*. 2017;25(1):43–56. <https://doi.org/10.1016/j.cmet.2016.09.018>.
 29. Barrea L, Pugliese G, Frias-Toral E, El Ghoch M, Castellucci B, Chapela SP, Carignano MLA, Laudisio D, Savastano S, Colao A, Muscogiuri G. Coffee consumption, health benefits and side effects: a narrative review and update for dietitians and nutritionists. *Crit Rev Food Sci Nutr*. 2023;63(9):1238–61. <https://doi.org/10.1080/10408398.2021.1963207>.
 30. Schubert MM, Irwin C, Seay RF, Clarke HE, Allegro D, Desbrow B. Caffeine, coffee, and appetite control: a review. *Int J Food Sci Nutr*. 2017;68(8):901–12. <https://doi.org/10.1080/09637486.2017.1320537>.
 31. Sirotkin AV, Kolesárová A. The anti-obesity and health-promoting effects of tea and coffee. *Physiol Res*. 2021;70(2):161–8. <https://doi.org/10.33549/physiolres.934674>.
 32. Lynch DH, Rushing BR, Pathmasiri W, McRitchie S, Batchek DJ, Petersen CL, Gross DC, Sumner SCJ, Batis JA. Baseline serum biomarkers predict response to a weight loss intervention in older adults with obesity: a pilot study. *Metabolites*. 2023;13(7). <https://doi.org/10.3390/metabo13070853>.
 33. Qu W, Chen Z, Hu X, Zou T, Huang Y, Zhang Y, Hu Y, Tian S, Wan J, Liao R, Bai L, Xue J, Ding Y, Hu M, Zhang XJ, Zhang X, Zhao J, Cheng X, She ZG, Li H. Profound perturbation in the Metabolome of a canine obesity and metabolic disorder model. *Front Endocrinol (Lausanne)*. 2022;13849060. <https://doi.org/10.3389/fendo.2022.849060>.
 34. Harris K, Desai N, Gupta M, Xue X, Chatterjee PK, Rochelson B, Metz CN. The effects of prenatal metformin on obesogenic diet-induced alterations in maternal and fetal fatty acid metabolism. *Nutr Metab (Lond)*. 2016;13(1):55. <https://doi.org/10.1186/s12986-016-0115-9>.
 35. Alves A, Bassot A, Bulteau AL, Pirola L, Morio B. Glycine metabolism and its alterations in obesity and metabolic diseases. *Nutrients*. 2019;11(6). <https://doi.org/10.3390/nu11061356>.
 36. Alsharari ZD, Leander K, Sjögren P, Carlsson A, Cederholm T, de Faire U, Hellenius ML, Marklund M, Risérus U. Association between carbohydrate intake and fatty acids in the de novo lipogenic pathway in serum phospholipids and adipose tissue in a population of Swedish men. *Eur J Nutr*. 2020;59(5):2089–97. <https://doi.org/10.1007/s00394-019-02058-6>.
 37. Jewesson PJ, Ensom RJ. Influence of body fat on the volume of distribution of theophylline. *Ther Drug Monit*. 1985;7(2):197–201. <https://doi.org/10.1097/00007691-198506000-00010>.
 38. Cranmer-Byng MM, Liddle DM, De Boer AA, Monk JM, Robinson LE. Pro-inflammatory effects of arachidonic acid in a lipopolysaccharide-induced inflammatory microenvironment in 3T3-L1 adipocytes in vitro. *Appl Physiol Nutr Metab*. 2015;40(2):142–54. <https://doi.org/10.1139/apnm-2014-0022>.
 39. Iyer A, Fairlie DP, Brown L. Lysine acetylation in obesity, diabetes and metabolic disease. *Immunol Cell Biol*. 2012;90(1):39–46. <https://doi.org/10.1038/icb.2011.99>.
 40. Asahi R, Tanaka K, Fujimi TJ, Kanzawa N, Nakajima S. Proline decreases the suppressive effect of histidine on Food Intake and Fat Accumulation. *J Nutr Sci Vitaminol (Tokyo)*. 2016;62(4):277–80. <https://doi.org/10.3177/jnsv.62.277>.
 41. Qian L, Fan Y, Gao F, Zhao B, Yan B, Wang W, Yang J, Ma X. Genetically determined levels of serum metabolites and risk of Neuroticism: a mendelian randomization study. *Int J Neuropsychopharmacol*. 2021;24(1):32–9. <https://doi.org/10.1093/ijnp/yyaa062>.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.